Reactions of Glycosyl Fluorides. Synthesis of O-, S-, and N-Glycosides

K. C. Nicolaou,* Alexander Chucholowski, Roland E. Dolle, and Jared L. Randall

Department of Chemistry, University of Pennsylvania, Philadelphia, Pennsylvania 19104, U.S.A.

Glycosyl fluorides react with a variety of *O-*, *S-*, and *N-*nucleophiles to afford the corresponding glycosides in good to excellent yields.

The high abundance and biological importance of carbohydrate-containing natural products, such as C-glycosides, lipopolysaccharides, glycoproteins, and nucleotides dictate new synthetic technology for their construction. In previous papers we described reactions of glycosyl fluorides leading to C-glycosides¹ and oligosaccharides.^{2,3} In this communication we report the utilization of these versatile intermediates in the synthesis of novel O-, S-, and N-glycosides, equation (1),†‡ all compounds of potential biological importance. Table 1 demonstrates the scope of this methodology with the preparation of glycosyl esters (entries 1,2,14), 'double' glycosides (entry 3), thioglycosides (entries 11,12), glycosyl peroxides (entry 6), glycosyl azides (entry 7), and aminoglycosides (entries 8—10). In most cases BF₃·Et₂O was found to be an effective catalyst for the coupling reaction. Amines were coupled to the carbohydrate residue after activation with AlMe₃ or by the action of MgBr₂·Et₂O, which presumably leads to the more active glycosyl bromide as an intermediate.1 The facile coupling of glycosyl fluorides to bases such as 6-azauridine via its disilyl ether after activation with SnCl₄ (entry 13)⁴ offers promising applications of the present technology in the construction of nucleotides, whereas the entry into glycosyl phosphate esters (entry 4) may provide the first stage for stereospecific enzyme-induced glycosidations.⁵ It is expected that the present methodology will find widespread applications in the synthesis of carbohydratecontaining naturally occurring and other synthetic molecules.

‡ ¹H N.m.r. data (250 MHz, CDCl₃, Me₄Si) (**2a**) δ 7.50—7.10 (m, 30H, Ph), 6.42 (d, J 4.5 Hz, 1-H), 4.97—4.52 (m, 8H, OC H_2 Ph), 3.94—3.68 (m, 6H, CHO, CH₂O), 3.89 and 3.61 (doublets, J 15.0 Hz, 2H each, NC H_2 Ph), 3.44 (dd, J 8.1, 6.9 Hz, 1H, CHCO₂), 1.84—1.45 (m, 3H, C H_2 CHMe₂), 0.80 and 0.61 (doublets, J 7.5 Hz, 3H each, CMe₂); (**2d**) 7.50—7.20 (m, 20H, Ph), 5.00—4.55 (m, 8H, C H_2 Ph), 3.93 (d, J 8.0 Hz, 1-H), 3.80—3.35 (m, 6H, CHO, CH₂O), 3.02—2.70 (m, 8H, morpholine); (**4e**) 7.40 (m, 5H, Ph), 5.90 (d, J 7.5 Hz, 1H, NH), 5.12 (m, 5H, 2-H, 3-H, 4-H, OC H_2 Ph), 4.59 (m, 1H, CHCO₂Et), 4.48 (d, J 8.0 Hz, 1-H), 4.20 (m, 4H, 6-H, OC H_2 Me), 3.63 (m, 1H, 5-H), 3.25 (dd, J 12.0, 4.5 Hz, 1H, SCH₂), 3.05 (dd, J 12.0, 5.0 Hz, 1H, SCH₂), 2.12, 2.08, 2.07, 2.05 (singlets, 3H each, OAc), 1.25 (t, J 5.0 Hz, 3H, OCH₂Me); (**4f**) 9.88 (br.s, 1H, NH), 7.51 (s, 1H, CH=N), 5.89 (d, J 7.5 Hz, 1H, 1-H), 5.69, 5.35, 5.15 (dd's, J 7.5 Hz, 1H each, 2-H, 3-H, 4-H), 4.20 (m, 2H, 6-H), 3.94 (m, 1H, 5-H), 2.14, 2.12, 1.99, 1.94 (singlets, 3H each, OAc).

PhCH₂O OCH₂Ph MeCO₂ O₂CMe

(1)

PhCH₂O OCH₂Ph MeCO₂ O₂CMe

(3)

PhCH₂O OCH₂Ph MeCO₂ O₂CMe

(3)

PhCH₂O OCH₂Ph MeCO₂ O₂CMe

(4)

$$\alpha$$
; R = O₂C OCH₂Ph OCH₂Ph

 α ; R = O₂C OCH₂Ph

 α ; R = O₂C OCH₂Ph

 α ; R = O₂C OCH₂Ph

 α ; R = OCH₂Ph

$$Me_3SiO \xrightarrow{N \\ N \\ N} N$$

(6) X = F(7) X = R (OCOMe)

[†] New compounds exhibited satisfactory spectroscopic and analytical data. Yields refer to pure isolated (flash column chromatographysilica) products.

Table 1. Synthesis of O-, S-, and N-glycosides.

Entry	Reagents (equiv.) and conditions	R	Yield ratio α:β) ^f
Substrate (1) $a,b,c \rightarrow \text{product}$ (2).			
1	MeCO ₂ H (4), BF ₃ ·Et ₂ O (0.5), CH ₂ Cl ₂ , 4Å MS, ^d $0\rightarrow$ 25 °C	Me	97 (ca. 3 : 2)
2	$RH(2)$, $BF_3 \cdot Et_2O(0.3)$, CH_2Cl_2 , 4Å MS , $0^{\circ}C$	(a)	76 (10:1)
3	$RH(1)$, $BF_3 \cdot Et_2O(0.3)$, CH_2Cl_2 , $0 \rightarrow 25 ^{\circ}C$	(b)	65e
4	(PhCH2O)2P(O)OSnBun3(2), BF3·Et2O(2),		
	$Et_2O, 0 \rightarrow 25 ^{\circ}C$	$O(O)P(OCH_2Ph)_2$	50 (10:1)
5	$RH(1)$, $BF_3 \cdot Et_2O(0.5)$, 4Å MS , $-15 \rightarrow 0$ °C	(c)	51 (2:1)
6	$Bu^{t}O_{2}H(2), BF_{3}\cdot Et_{2}O, (0.2), CH_{2}Cl_{2},$		
	4Å MS, −15→0 °C	O_2Bu^t	91 (3:1)
7	Me_3SiN_3 (2.5), $BF_3 \cdot Et_2O$ (0.5), CH_2Cl_2 ,		
	0→25 °C	N_3	90 (10:1)
8	$RH (1.5), MgBr_2 \cdot Et_2O (5), CH_2Cl_2, 25 ^{\circ}C$	(d)	90 (1:10)
9	$H_2NCH_2=CH_2(1)$, $AlMe_3(1)$, CH_2Cl_2 , 25 °C	NHCN ₂ CH=CH ₂	95 (2:1)
10	$H_2NPh(2)$, AlMe ₃ (2), CH_2Cl_2 , 25 °C	NHPh	65 (1:1)
Substrate (3) $a,b \rightarrow product$ (4).			
11	$RH(1.5), BF_3 \cdot Et_2O(0.1), CH_2Cl_2, 25 ^{\circ}C$	(e)	80 (1:20)
12	PhSH (2), BF ₃ ·Et ₂ O (0.1), CH ₂ Cl ₂ , 25 °C	SPh	95 (1:20)
13	(5) (1.5), SnCl ₄ (0.2), CH ₂ Cl ₂ , 25 °C	(f)	76 (1:20)
	2 2	(-)	/ ((1 . = 0)
Substrate $(6)^{a,b,c} \rightarrow \text{product } (7).^b$			
14	$MeCO_2H(4)$, $BF_3 \cdot Et_2O(0.5)$, CH_2Cl_2 , $4Å$ MS,		
	0→25 °C	OCOMe	83 (3:2)

^a Prepared from the corresponding phenylthioglycoside and N-bromosuccinimide–diethylamino sulphur trifluoride (ref. 2); ^b structure determined by spectroscopic methods; ^c $\alpha:\beta$ mixture ca. 1:1; ^d MS = molecular sieve; ^e ratio not determined; ^f ratio was determined by ¹H n.m.r.

Financial support for this work from the National Institutes of Health (U.S.A.) and Merck Sharp & Dohme (U.S.A.) is gratefully acknowledged.

Received, 2nd May 1984; Com. 613

References

 K. C. Nicolaou, R. E. Dolle, A. Chucholowski, and J. L. Randall, J. Chem. Soc., Chem. Commun., 1984, preceding communication.

- K. C. Nicolaou, R. E. Dolle, D. P. Papahatjis, and J. L. Randall, J. Am. Chem. Soc., 1984, 106, 4189.
- 3 K. C. Nicolaou, S. P. Seitz, and D. P. Papahatjis, J. Am. Chem. Soc., 1983, 105, 2430.
- 4 U. Nedballa and H. Vorbruggen, J. Org. Chem., 1974, 39, 3654.
- 5 N. K. Kochetkov, in 'Bacterial Lipopolysaccharides,' ACS Symposium Series 231, 1983, Ch. 4, p. 65; J. F. Kennedy and C. A. White, in 'Bioactive Carbohydrates,' Wiley, New York, 1983, Ch. 5, p. 88.